



An unsupervised machine learning method for discovering patient clusters based on genetic signatures

Christian Lopez ^a, Scott Tucker ^{b, c}, Tarik Salameh ^b, Conrad Tucker ^{a, d, e, *}

^a Industrial and Manufacturing Engineering, The Pennsylvania State University, University Park, PA 16802, USA

^b Hershey College of Medicine, The Pennsylvania State University, Hershey, PA, 17033, USA

^c Engineering Science and Mechanics, The Pennsylvania State University, University Park, PA 16802, USA

^d Engineering Design Technology and Professional Programs, The Pennsylvania State University, University Park, PA 16802, USA

^e Computer Science and Engineering, The Pennsylvania State University, University Park, PA 16802, USA

ARTICLE INFO

Article history:

Received

Received in revised form

Accepted

Available online

ABSTRACT

Introduction: Many chronic disorders have genomic etiology, disease progression, clinical presentation, and response to treatment that vary on a patient-to-patient basis. Such variability creates a need to identify characteristics within patient populations that have clinically relevant predictive value in order to advance personalized medicine. Unsupervised machine learning methods are suitable to address this type of problem, in which no *a priori* class label information is available to guide this search. However, it is challenging for existing methods to identify cluster memberships that are not just a result of natural sampling variation. Moreover, most of the current methods require researchers to provide specific input parameters *a priori*.

Method: This work presents an unsupervised machine learning method to cluster patients based on their genomic makeup without providing input parameters *a priori*. The method implements internal validity metrics to algorithmically identify the number of clusters, as well as statistical analyses to test for the significance of the results. Furthermore, the method takes advantage of the high degree of linkage disequilibrium between single nucleotide polymorphisms. Finally, a gene pathway analysis is performed to identify potential relationships between the clusters in the context of known biological knowledge.

Datasets and Results: The method is tested with a cluster validation and a genomic dataset previously used in the literature. Benchmark results indicate that the proposed method provides the greatest performance out of the methods tested. Furthermore, the method is implemented on a sample genome-wide study dataset of 191 multiple sclerosis patients. The results indicate that the method was able to identify genetically distinct patient clusters without the need to select parameters *a priori*. Additionally, variants identified as significantly different between clusters are shown to be enriched for protein-protein interactions, especially in immune processes and cell adhesion pathways, via Gene Ontology term analysis.

Conclusion: Once links are drawn between clusters and clinically relevant outcomes, Immunochip data can be used to classify high-risk and newly diagnosed chronic disease patients into known clusters for predictive value. Further investigation can extend beyond pathway analysis to evaluate these clusters for clinical significance of genetically related characteristics such as age of onset, disease course, heritability, and response to treatment.

Keywords:

Unsupervised machine learning

Clustering analysis

Genomic similarity

Multiple Sclerosis

* Corresponding author. 213 N Hammond Building, University Park, PA 16802, USA.

E-mail address: ctucker4@psu.edu (C.S. Tucker).

1. Introduction

With advancements in genome-wide association study (GWAS) techniques and the advent of low cost genotyping arrays, researchers have developed a significant interest in applying Machine Learning (ML) methods to mine knowledge from patients' genomic makeup [1,2]. This knowledge has allowed researchers to improve gene annotation and discover relationships between genes and certain biological phenomena [3,4].

The fields of personalized and stratified medicine benefit greatly from ML. For example, many cases in the field of pharmacogenetics have identified genetic variants with clinically actionable impacts on drug response and metabolism [5,6]. Moreover, many chronic disorders (e.g., asthma, diabetes, Crohn's disease) have genomic etiology, clinical presentation, and response to treatment that vary on a patient-to-patient basis. Such variability reveals a need to identify characteristics within patient populations that have clinically relevant insights. For example, Multiple Sclerosis (MS) is a chronic inflammatory disorder in which progressive autoimmune demyelination and neuron loss occur in the central nervous system. MS varies from patient-to-patient in genomic etiology, disease progression, clinical presentation, and response to treatment. Hence, MS patients, like other chronic autoimmune patients, could benefit from ML methods that advance personalized medicine.

Machine learning methods are commonly classified into *supervised* and *unsupervised* methods. Supervised methods, such as Support Vector Machines [7] and Random Forests [8,9], have been extensively used in the field of bioinformatics. These methods classify new objects to a determinate set of discrete class labels while minimizing an empirical loss function (e.g., mean square error). However, supervised methods require the use of a training set that contains *a priori* information of several objects' class labels. In contrast, unsupervised methods do not require a training set that contains *a priori* information of objects' class labels as input. Unsupervised methods are able to detect potentially interesting and new cluster structures in a dataset. Moreover, they can be implemented when class label data is unavailable. Hence, if the objective of a study is to discover the class labels that best describe a set of data, unsupervised machine learning should be implemented in place of supervised methods [2]. However, it is challenging for existing unsupervised ML methods to identify object memberships that are due to the underlying cluster structures in the dataset, rather

than the results of natural sampling variation [10]. Moreover, most current methods require researchers to provide certain input parameters *a priori* (e.g., number of clusters in the dataset), which can limit their applicability.

In light of the limitations of existing methods and the need to advance personalized medicine, an unsupervised machine learning method to cluster patients based on their genomic similarity is presented. The method integrates statistical analysis that accounts for *family-wise-error* rate, allowing the method to identify clusters resulting from the underlying structure of the data and not just due to random chance. Moreover, the method takes advantage of the high degree of linkage disequilibrium between Single Nucleotide Polymorphisms (SNP) by pruning correlated nearby SNPs, which helps reduce redundant variants in the dataset. Finally, a gene pathway analysis shows the potential relationships between the clusters in the context of known biological knowledge. The proposed method is capable of clustering patients based on their genomic similarity without *a priori* information. Moreover, it is capable of identifying the significant variants (i.e., SNPs) between patient sub-groups within a cohort with a common disorder. Successfully identifying distinct genetic subtypes of patients within genomic datasets demonstrates the potential of this method to advance personalized medicine of complex diseases with heritable components, especially autoimmune disorders which have many shared susceptibility loci [11].

2. Literature review

In the last decade, the field of bioinformatics has seen a significant number of publications implementing unsupervised machine learning methods, such as clustering algorithms [12-14]. Clustering algorithms partition data objects (e.g., genes, patients) into groups (i.e., clusters), with the objective of exploring the underlying structure on a dataset [15]. In the medical field, these algorithms have been implemented to identify sets of co-expressed genes [16], compare patients' prognostic performance [17], cluster patients based on their medical records [18], and identify subgroups of patients based on their symptoms and other variables [19].

In previous work, genomic stratification of patients (i.e., stratified medicine) has been able to match specific therapy recommendations to genetic subpopulations by predicting therapeutic response [5,6]. However, most of these studies implemented class label data (i.e., response to treatment) to

cluster patients. In clinical datasets, class label information is not widely available for convenient patient clustering. Unsupervised machine learning methods can be used in such cases to identify clusters within the dataset. Further investigation of genetic subgroups within a cohort of patients can offer a better clinical prediction of age of onset, disease course, heritability, and response to therapy, leading to improved outcomes [20].

2.1. Hierarchical clustering algorithms.

Agglomerative hierarchical clustering algorithms are one of the most frequently used algorithms in the biomedical field [21,22]. Researchers have found that hierarchical clustering algorithms tend to perform better than other algorithms (e.g., k-means, partitioning around Medoids, Markov clustering) when tested on multiple biomedical datasets [23]. The objective of any agglomerative hierarchical clustering algorithm is to cluster a set of n objects (e.g., patients, genes) based on an $n \times n$ similarity matrix. These clustering algorithms have grown in popularity due to their capability to simultaneously discover several layers of clustering structure, and visualize these layers via tree diagrams (i.e., dendrogram) [10]. Even though these algorithms allow for easy visualization, they still require preselecting a similarity height cut-off value in order to identify the final number of clusters. In other words, it still requires researchers to know *a priori* the number of cluster in the dataset.

Agglomerative hierarchical clustering algorithms can be implemented with different linkage methods. For example, Ahmad *et al.* (2016) [17] implemented the Ward's linkage method to compare patients' prognostics performance; while Hamid *et al.* (2010) [19] implemented the Complete linkage method to identify unknown sub-group of patients. Unfortunately, depending on the underlying structure of the data, different clustering results can be obtained by implementing different linkage methods. Ultsch and Löttsch (2017) [24] demonstrated that neither the Single nor Ward's linkage methods provided similar clustering results when tested with the Fundamental Clustering Problem Suite (FCPS) datasets [25]. Their results reveal that these linkage methods were able to correctly cluster all the objects in only a subset of the FCPS datasets. Similarly, Clifford *et al.* (2011) [26] discovered that while testing multiple simulated GWAS datasets, the linkage methods of Median and Centroid were the only ones to consistently be outperformed by the Single, Complete, Average, Ward's, and McQuitty methods. In light of these, Ultsch and

Löttsch (2017) [24] proposed the use of emergent self-organizing map to visualize clustering of high-dimensional biomedical data into two-dimensional space. Even though, their method allowed for better visualization, it still required preselecting the number of clusters as well as other parameters to perform correctly (e.g., toroid grid size) [24].

2.2. Parameter selection in clustering algorithms.

In order to avoid preselecting input parameters *a priori* (e.g., the number of clusters), researchers have implemented cluster validation metrics. For example, Clifford *et al.* (2011) [26] proposed a method that aimed to capture the clustering outcome of multiple combinations of linkage method and similarity metric based on the Silhouette index [27]. The Silhouette index was used to rank the results of the clustering combinations, and select the best cluster set (i.e., cluster set with largest average Silhouette index). Similarly, Pagnuco *et al.* (2017) [16] presented a method that implemented several linkage methods and implemented modified versions of the Silhouette and Dunn indices [28] to select the final clustering results. Both the Silhouette and Dunn indices served as internal cluster validation metrics (i.e., no external information needed) to guide the selection of the final cluster set. However, the Silhouette index has been shown to have a stronger correlation with external cluster validation metrics, such as the Rand Index, than the Dun index [28,30].

The methods of Clifford *et al.* (2011) and Pagnuco *et al.* (2017) did not require selecting the number of clusters *a priori* due to the internal cluster validation metrics implemented. These metrics allow for algorithmic selection of the number of clusters. Nonetheless, the computational complexity of testing all potential clusters increases linearly with the number of objects in the dataset. Other studies have implemented model-based clustering methods to overcome these limitations. For example, Sakellariou *et al.* (2012) [29] implemented an Affinity Propagation [30] algorithm to identify relevant genes in microarray datasets. Shen *et al.* (2009) [31] implemented an Expectation-Maximization algorithm [32] to cluster genes based on an integration of multiple genomic profiling datasets. However, models based methods make underlying assumptions that might not be applicable in certain datasets [33].

Recently, Khakabimamaghani and Ester (2016) [34] presented a Bayesian biclustering method to identify clusters of patients. They benchmarked their method against the multiplicative Non-negative Matrix Factorization (NMF) algorithm proposed by Lee

and Seung (2001) [35]. Their results revealed that their Bayesian biclustering method was more effective in patient stratification than the NMF. While this Bayesian biclustering method did not require selecting the number of clusters *a priori*, it did require selecting parameters for prior probability distributions. The capability of biclustering algorithms to discover related gene sets under different experimental conditions, have made them popular within the bioinformatics community [36]. One of the first works in this area was presented by Cheng and Church (2000) [37]. They proposed an iterative greedy search biclustering algorithm to cluster gene expression data. Even though their method did not require selecting the number of clusters *a priori*, it did require the selection of hyperparameters (e.g., maximum acceptable error).

2.3. Statistical significance of clustering results.

Even though the methods of Clifford *et al.* (2011) and Pagnuco *et al.* (2017) aimed to find the optimal clustering outcome from multiple algorithms, which resembled the consensus clustering approach (i.e., approach in which a solution is identified by validating multiple outcomes) [38], their methods did not account for possible clustering memberships arising due to random variation. Whether identified clusters memberships are due to underlying cluster structures in the data or are just a result of the natural sampling variation, is a critical and challenging question that needs to be addressed when clustering high-dimensional data [10]. To address this question, Suzuki and Shimodaira (2013) [39] presented the *pvclust* R package, which calculates probability values for each cluster using nonparametric bootstrap resampling techniques. Even though *pvclust* allows for parallelized computing, it requires significant time (i.e., 480 mins) when implemented in genomic datasets. This is due to the large number of resampling iterations (i.e., 10,000) required to reduce the error rate [39]. In contrast, Ahmad *et al.* (2016) [17] applied a non-parametric analysis of variance (ANOVA) Kruskal-Wallis test to compare the clusters within a hierarchical clustering method. Similarly, Bushel *et al.* (2002) [40] implemented a single gene parametric ANOVA test to assess the effects of genes on hierarchical clustering results. Recently, Kimes *et al.* (2017) [10] proposed a method based on a Monte Carlo approach to test the statistical significance of hierarchical clustering results while controlling for *family-wise-error* rate. However, *family-wise-error* rate can also be controlled while applying repetitive statistical tests by implementing a Bonferroni correction [41].

2.4. Integrating domain knowledge into clustering algorithms

Other frequently used clustering algorithms in the bioinformatics field are k-means and fuzzy c-means. However, these algorithms require initial random assignments of the clusters, which can produce inconsistent results [26]. Hence, they might fail to converge to the same results, even after multiple initiations using the same dataset [21]. In light of these limitations, Tari *et al.* (2009) [21] proposed the “*GO Fuzzy c-means*” clustering algorithm. Their method resembles the fuzzy C-mean algorithms [42] and implements Gene Ontology annotation [43] as biological domain knowledge to guide the clustering procedure. Even though this method assigned genes to multiple clusters, which could have improved the biological relevance of the results, it was not capable of discriminating the cluster memberships that were assigned due to random chance. While the algorithm parameters selected in this study might have been reasonable for the dataset analyzed, the authors highlighted that future studies would need to experimentally determine these parameters. Similarly, Khakabimamaghani and Ester (2016) [34] integrated domain knowledge via the selection of parameters for prior probability distributions. However, their results reveal that the selection of these parameters had a direct impact on their clustering results. When analyzing the effects of priors, the authors indicate that “*final selected priors favor better sample clustering over better gene clustering*” [34]. These findings reveal that the parameters need to be carefully selected since they can bias their method towards better sample clustering rather than better gene clustering results.

Researchers can implicitly integrate domain knowledge to their methods by judiciously selecting the input data of their algorithms [2]. Genomic datasets may include relevant features as well as correlated and non-informative features. The presence of correlated and non-informative features might obscure relevant patterns and prevent an algorithm from discovering the underlying cluster structure of a dataset [19]. Genomic data is generally high-dimensional because the number of features is frequently greater than the number of samples. Additionally, genetic variants are commonly correlated with other variants in close proximity on DNA. Therefore, when clustering genomic data, it is important to prune non-informative and correlated features [2,9].

Highly correlated SNPs are said to be in Linkage Disequilibrium (LD). This characteristic makes it challenging for

unsupervised ML algorithms to discover relevant cluster structures in the dataset. GWA studies present significant associations as tag SNPs, implying a true causal SNP can be found within the LD block of a tagged location [11]. LD pruning refers to removing highly correlated SNPs within LD blocks. For example, Yazdani *et al.* (2016) [44] identified a subset of informative SNPs based on a correlation coefficient. Similarly, Goldstein *et al.* (2010) [9], implemented several correlation coefficient cut-off values (e.g., 0.99, 0.9, 0.8, 0.5) to remove SNPs with high LD. They achieved this by using the toolsets for Whole-Genome Association and Population-Based Linkage Analyses (PLINK) [45], resulting in a reduction of up to 76% of the original dataset. This reduction decreased the computational complexity of their method [9]. However, researchers have not agreed yet on a standard correlation coefficient cut-off value that can be applied to every genomic dataset to reduce complexity without incurring in significant information loss.

Table 1. Summary of current methods

Papers	LD Pruning	Automatic selection of k^*	Statistical tests performed	No selection of parameters required †
[9,11,19, 21]	X			
[40]	X		X	
[16,26,29, 31,34, 35,37]		X		
[10, 17, 39, 46]			X	
<i>This work</i>	X	X	X	X

* k is the parameter defining the number of cluster in the dataset.

† No parameters or hyperparameters are required to be known or selected *a priori* by researchers (e.g., prior probability, toroid grid size).

Table 1 shows a summary of the current clustering methods in the field of bioinformatics applied to genomics data. It can be shown that multiple methods prune the SNPs of their datasets based on the degree of LD between nearby SNPs. This is done in order to guide their clustering search and remove potentially non-informative features. However, the vast majority of existing methods still require preselecting the number of clusters and other parameters *a priori* (e.g., prior probability distributions, toroid grid size). Moreover, the current methods do not commonly implement statistical analysis to test for the significance of their results, or to account for possible *family-wise-error* rates.

In light of the aforementioned limitations, an unsupervised machine learning method is presented in this work that seeks to identify sub-groups within cohorts of patients afflicted with the same disease. This is done by clustering patients based on their genomic similarity without the need of *a priori* input parameters.

The method presented in this work takes advantage of LD between SNPs by pruning correlated SNPs. In addition, it automatically selects the number of clusters by implementing an internal validation metric. The method ensembles the clustering outcomes of multiple linkage methods via a majority vote approach. Subsequently, it tests for statistical significance among results while accounting for *family-wise-error* rate. Finally, a gene pathway analysis is performed to support the potential medical significance of the results.

3. Method

An unsupervised machine learning method is presented that does not require selection of input parameters *a priori*. The method can help identify patient cluster structures within genomic data and potentially discover valuable differences between them. This knowledge can be used to advance personalized medicine of complex diseases with heritable components, especially autoimmune disorders which have many susceptibility loci. Fig. 1 shows an outline of the method presented in this work.

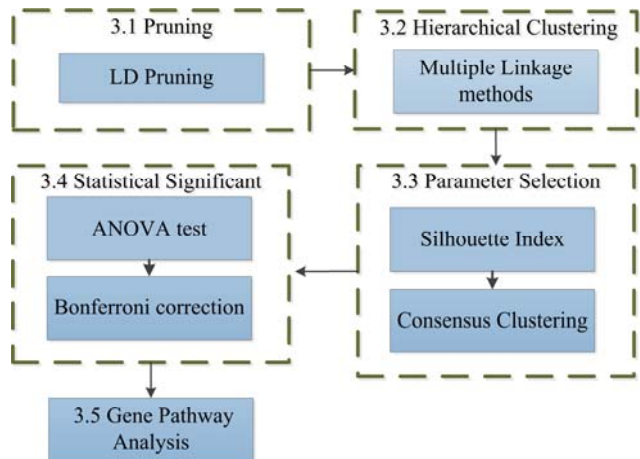


Figure 1. Outline Method

3.1 Linkage Disequilibrium Pruning

Pruning SNPs based on LD serves as a feature reduction step. Thus, in the proposed method, SNPs that are strongly correlated to other nearby SNPs are pruned, as previously done in the literature. The degree of LD between SNPs is assessed by calculating the correlation coefficients based on a sliding window method. In this method, cut-off values of (i) 0.999, (ii) 0.99, (iii) 0.9, (iv) 0.8 and (v) 0.5 are employed. Previous studies have shown these cut-off values provide a balance between error reduction and information loss [9]. Hence, five subsets of patients' genomic data containing different sets of SNPs (i.e.,

features) are generated. The subsets generated serve as input for the hierarchical clustering step.

3.2 Hierarchical Clustering

The objective of the unsupervised machine learning method presented in this work is to cluster patients based on their genomic similarity. Patients' genomic similarity can be evaluated using a wide range of distance metrics [26]. The selection of the appropriate distance metric is driven by the type of data under analysis (e.g., ratio, interval, ordinal, nominal or binary scale). For example, the Euclidian distance is appropriated for ratio or interval scale data, while the Manhattan distance for ordinal scale data [47].

Subsequently, the method presented in this work employs an agglomerative hierarchical clustering algorithm. Hierarchical clustering algorithms are frequently used with only one linkage method, which can limit their ability to identify underlying cluster structures in certain datasets [24]. Hence, in this work, multiple linkage methods are implemented. The linkage methods used in this work have been shown to consistently outperform other methods when tested with simulated GWAS datasets [26]. The cluster results obtained by implementing different linkage methods are ensemble in the subsequent steps. This ensemble takes advantage of the performance of multiple linkage methods. Moreover, it helps identify the underlying structure of the data, since the ensemble approach will favor cluster structures identified by the majority (i.e., via a majority vote approach) of the linkage methods. Specifically, the authors propose to implement:

- (i) *Single Linkage (or Minimum Linkage).*
- (ii) *Complete Linkage (or Maximum Linkage).*
- (iii) *Average Linkage (or Unweighted Pair Group Method with Arithmetic Mean, UPGMA).*
- (iv) *Ward's Linkage.*
- (v) *McQuitty Linkage (or Weighted Pair Group Method with Arithmetic Mean, WPGMA).*

3.3 Parameter Selection

Once the agglomerative hierarchical algorithm is implemented, the Silhouette index is employed as an internal validity metric. This index has been used in previous studies to rank the results of multiple clustering algorithms outcomes and guide the selection of final clusters [16],[26]. Nonetheless, in this method, the index

is used to select the number of clusters for all combinations of LD pruning data subsets (see section 3.1) and linkage methods (see section 3.2). The number of clusters that provides the largest average Silhouette index value in each of the combinations is selected.

The computational complexity of testing all possible numbers of clusters increases linearly as the number of objects in a dataset increases. This can be a challenge in datasets that contain a large number of objects, even with parallelized computing. In this work, an optimization approach is presented to identify the number of clusters that maximizes the average Silhouette index. The mathematical formulation of this optimization problem is as follows:

$$\text{Maximize } SI \quad (1)$$

$$SI = \frac{1}{k} \sum_{i=1}^k \left[\frac{1}{\eta_i} \sum_{x \in C_i} S(x) \right] \quad \forall i \in \mathbf{K}\{1, \dots, k\} \quad (2)$$

$$S(x) = \frac{b(x) - a(x)}{\max\{b(x), a(x)\}} \quad \forall x \in C_i \quad (3)$$

$$a(x) = \frac{1}{\eta_i - 1} \sum_{y \in C_i} d(x, y) \quad \forall x \neq y, x \text{ and } y \in C_i \quad (4)$$

$$b(i) = \min_{w \in \mathbf{K}, w \neq i} \left\{ \frac{1}{\eta_w} \sum_{g \in C_w} d(x, g) \right\} \quad \forall g \in C_w, w \in \mathbf{K} \quad (5)$$

$$1 < k \leq n \quad (6)$$

Where,

SI : is the average Silhouette index of the clusters set \mathbf{K}

\mathbf{K} : is the set of clusters obtained with the hierarchical clustering algorithm for a given number of k disjoint clusters

η_i : is the number of objects that belongs to cluster C_i , for $i \in$ set of clusters $\{\mathbf{K}\}$

$S(x)$: is the Silhouette of object x , for $x \in C_i$

$a(x)$: is the average similarity of object x with all other objects that belong to the same cluster of x (i.e., C_i)

$b(x)$: is the average similarity of object x with the objects from the nearest clusters C_w , for $w \in$ set of cluster $\{\mathbf{K}\}$, $i \neq w$.

Eq. (1) represents the objective function that needs to be maximized (i.e., the average Silhouette index). Eq. (2), shows the mathematical representation of the average Silhouette index, while Eq. (3) shows the silhouette of a given object x . Both Eq. (4) and (5) represent the elements that constitute the Silhouette index of a given object x [27]. Finally, Eq. (6) constrains the search for the number of clusters to be greater than 1 and less than the total number of objects n (i.e., the maximum number of clusters). Since the objective function is non-linear with respect to the parameter k (i.e., number of clusters), this optimization

problem needs to be solved with a non-linear optimization algorithm. In the literature, there are several algorithms suitable to solve this type of optimization problem [48]. Nonetheless, the method is not constrained to any specific optimization algorithm.

Once the number of clusters is identified in all datasets combination, the results are aggregated into a final cluster set via a majority vote approach. Table 2 shows an example of this consensus clustering approach in which patient i is assigned to the final cluster 1 since the majority of the cluster results assigned that patient to that given cluster. Similarly, patient n is assigned to the final cluster 2, since the majority of clusters assigned this patient to this cluster.

Table 2. Example of consensus clustering

No.	<i>LD Pruning</i>	<i>Linkage Method</i>	<i>Patient i cluster</i>		<i>Patient n cluster</i>
1	1	Single	1	...	2
2	0.99	Single	1		2
3	0.90	Single	1		1
4	0.8	Single	1		2
5	0.5	Single	2		1
...					...
24	0.8	McQuitty	2		2
25	0.5	McQuitty	1		2
Final Cluster			1		2

3.4 Statistical Significance

After the final patient clusters are discovered, a single SNP ANOVA test is performed to reveal the SNPs that are statistically significantly different between the clusters of patients. This step helps validate that the clusters generated are different by at least one associated SNP. To account for *family-wise-error-rate* a Bonferroni correction is applied by dividing the alpha value by the number of tested SNPs. In the case that no SNPs are found to be statistically significantly different, it can be concluded that the resulting patients' clusters might have arisen due to random chance.

3.5 Gene Pathway Analysis

The set of SNPs significantly associated with differences between patient clusters can be explored via Gene Ontology (GO) enrichment and mutational burden on molecular pathways. By assigning each SNP to a gene and performing a gene network analysis, (e.g., via STRING-DB software [49]), visualizations of

gene networks and estimations of significant enrichment along GO terms could provide evidence for potential biological significance. The significance is assessed by comparing the number of evidence-based relationships between selected genes to the number expected in a set of randomly selected genes. If an enrichment is established, the related genes are examined by their molecular function, biological relevance, and known associations to the disease from GWA studies. While pathway analysis does not provide the rigor of direct experiment or clinical trial, it remains valuable in determining whether selected genes are functionally relevant to the disease studied, as opposed to being a function of other factors such as ethnicity.

4. Application

The performance of the proposed method is first tested on the datasets presented in the Fundamental Clustering Problem Suite (FCPS) [25]. The FCPS contains 10 different datasets designed to assess the performance of unsupervised machine learning algorithms on particular clustering challenges (e.g., outliers, undefined cluster boundaries). The ground truth data of cluster membership are used to test the performance of the method in identifying clusters resulting from the underlying structures in the data and not just from random variation. To measure this performance, the Rand index [50] validation metric is employed. Moreover, the performances of other existing methods in the literature are benchmarked with the same datasets. All the benchmark analyses were performed on a 12 Core i7 3.4 GHz Intel™ computer with 62.8 GB of RAM and Ubuntu 16.04 LTS. The benchmark methods were implemented in R v.3.4 [51] with the used of the packages *mclust* v.5.3 [52], *apcluster* v.1.4.4 [53], *DatabionicSwarm* v.0.9.8 [54,55], *NNLM* v.0.4.1 [35], and *biclust* v.1.2.0 [56].

Two genomic datasets are used to compare the performance of the proposed method against other state of the art methods in the literature. The benchmark methods include those that do not require providing the number of cluster *a priori*. I.e., Clifford *et al.* (2011) [26]: hierarchical clustering algorithm with silhouette index, Sakellariou *et al.* (2012) [29]: Affinity Propagation clustering algorithm, Shen *et al.* (2009) [31]: Expectation Maximization clustering algorithm, and Cheng and Church (2000) [37]: Iterative Greedy Search Bicustering algorithm. First, the microarray gene expression data of patients with lymphoblastic and acute myeloid leukemia from Golub *et al.* (1999) [57] was implemented. The dataset is publically available at the Broad Institute and has been previously used to test the

performance of clustering algorithms [23,58]. The dataset is composed of microarray gene expression data of 999 genes for 27 patients with acute lymphoblastic leukemia and 11 patients with acute myeloid leukemia.

Lastly, a dataset of patients diagnosed with Multiple Sclerosis (MS) is employed. DNA samples from 191 MS patients consented via the Pennsylvania State University PRIDE protocol at Hershey Medical Center were subjected to the Immunochip assay (Illumina). Allelic variations were measured at previously described susceptibility loci for multiple immune-mediated disorders [59,60]. The *Y* chromosome data were filtered out of the dataset to simplify comparisons in a predominantly female cohort. Mitochondrial markers were discarded for analysis as well. Genotype calling was done with Illumina *GenomeStudio* v.2011.1 (www.illumina.com), and genotype markers were excluded if their *GenTrain* score was less than 0.8, or if their call rate across the cohort was less than 0.99. Finally, the MS dataset was filtered such that only variants within coding regions (i.e., exons), were considered. Therefore, the MS dataset was composed of 191 patients and 25,482 SNPs.

With the MS dataset, a 10-fold cross-validation analysis was performed with the objective to test the performance of the proposed and the benchmark methods, as well as to provide evidence regarding their propensity of overfitting genomic datasets. In this cross-validation approach, the MS dataset was randomly partitioned into 10 subsets. Subsequently, the methods were used to cluster the patients within these subsets. The clustering results obtained from the 10 subsets were compared to those from the complete dataset. The agreement between the clusters generated with the complete MS dataset and the 10-fold subsets is assessed with the Rand index metric. A match between the clustering results (e.g., average Rand index of 1) will indicate that the method was not overfitting the MS dataset, thus, providing arguments of its generalizability. Moreover, it will support that the method was identifying clusters due to underlying structures in the data and not just due to random variations. Finally, the groups of SNPs identified by the proposed method to achieve statistical significance between clusters generated were examined via gene pathway analysis.

4.1 Linkage Disequilibrium Pruning

For the MS dataset, the pruning of SNPs with a high LD was done based on the correlation-coefficient cut-off values found in the literature, as proposed in section 3.1. LD pruning was

performed using the widely used genotype analysis toolset for Whole-Genome Association and Population-Based Linkage Analyses (i.e., PLINK) [45]. This pruning resulted in a reduction of the original dataset as presented in Table 3. These percentages of SNPs removed are consistent with the results found in previous studies.

Table 3. LD Pruning summary

R ² cut-off value	Number of SNPs retained	Percentage of SNPs removed
0.50	5,460	78.57%
0.80	6,849	73.12%
0.90	7,421	70.88%
0.99	8,666	65.99%
0.999	8,691	65.89%

4.2 Hierarchical Clustering

The FCPS and Golub *et al.* (1999) [57] datasets contain features that are in ratio scale. Hence, to measure the similarity between the objects in the datasets, the Euclidian distance is implemented. Genotype data can be ordinal or additive scale, depending on whether heterozygous SNPs are treated as a label or as a half-dosage. While additive models are more often used for GWA studies, in this work, ordinal scale was used to demonstrate flexibility in the described clustering method. Hence, the genomic similarity of MS patients based on different subsets of pruned data is evaluated using the Manhattan distance metric. The similarity calculations and the agglomerative hierarchical algorithm with multiple linkage methods were performed in R v.3.4 [51].

4.3 Parameter Selection

The selection of the number of clusters k that maximized the average Silhouette index was performed with a generalized simulation annealing algorithm. This algorithm was selected due to its underlying theory and proven performance in problems with non-linear objective functions [61,62]. The algorithm was implemented via the R package *GenSA* v.1.1.6 [63]. Nonetheless, other non-linear optimization algorithms or greedy heuristics can also be implemented. Once the number of clusters in every combination of LD pruned data and linkage method are selected, the clustering results are ensemble via a majority vote approach (see section 3.3).

4.4. Statistical Significance

After the final clusters have been selected based on the average Silhouette metric and consensus clustering approach the statistical significance of the results is evaluated. Clusters' median values for each of the p features in the MS dataset are evaluated via a single SNP non-parametric ANOVA Kruskal-Wallis test [46]. To account for *family-wise-error* rate, a Bonferroni correction is applied to the significance alpha level of 0.05 (i.e., Bonferroni correction = $0.05/p$, for $p=25,482$).

4.5. Gene Pathway Analysis

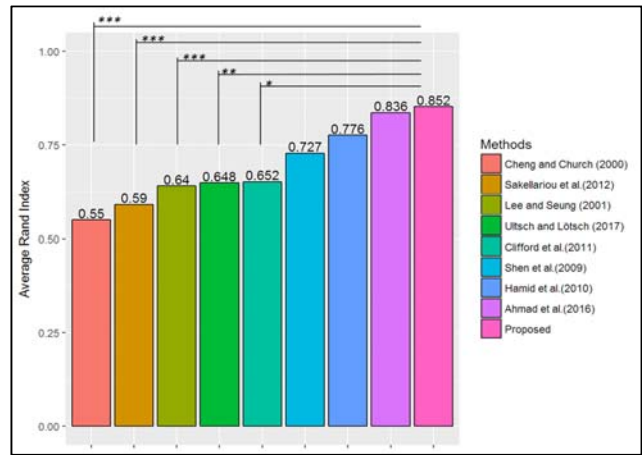
Gene variants that show statistical significance are further analyzed via a gene pathway analysis to explore their potential medical significance. Pathway analysis starts with generating a list of genes determined from the set of SNPs with strong evidence of significance between patient clusters. Inputting the gene set via the STRING-DB software algorithms [49] allows for convenient calculation of pathway enrichment hypothesis tests and visualization of the gene network. STRING-DB determines gene relationships by aggregating several databases into an evidence score. Experimental evidence comes from the BIND [64], GRID [65], HPRD [66], IntAct [67], MINT [68], and PID [69] databases. In addition, STRING-DB pulls from the curated databases KEGG [70], Gene Ontology [43], BioCarta [71], and Reactome [72]. Interaction frequency is tested for enrichment compared to expectation from a random sampling of genes, with p -values and false discovery rates reported for enrichment in specific cellular processes, defined by Gene Ontology references. After statistical testing is done, the gene network is used as a threshold for high confidence interaction and a k -means clustering algorithm is performed for visualization purposes (see Fig. 6).

5. Results

5.1. FCPS Benchmark results

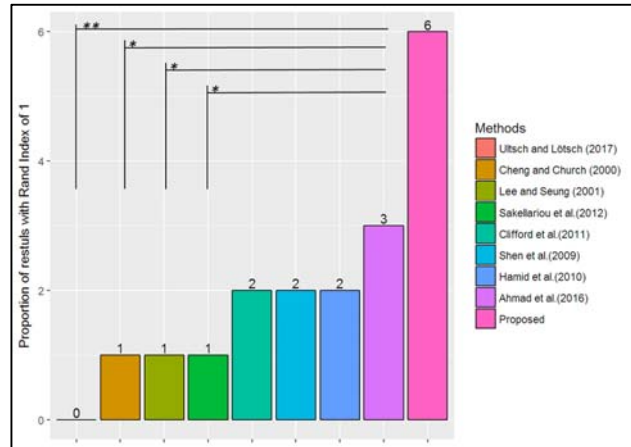
The majority of existing methods in the literature require the selection of parameters *a priori* (e.g., number of clusters, see Table 1). Hence, to benchmark with multiple methods, the number of clusters provided by the FCPS was used as input when testing these methods. Figure 2 shows the average Rand index obtained in the FCPS datasets by the method proposed in this work (i.e., *Proposed*) and the methods benchmarked. This plot shows that on average the proposed method outperformed other

methods, with an average Rand index of 0.852. The performance is statistically significantly greater than the results of the methods proposed by Cheng and Church (2000), Sakellariou *et al.* (2012), Lee and Seung (2001), Ultsch and Löttsch (2017), and Clifford *et al.* (2011). Even though these results indicate that, on average, the proposed method achieved the largest Rand index, there is not enough evidence to conclude that it was statically significantly greater than the Rand index achieved by the methods of Shen *et al.* (2009), Hamid *et al.* (2010), or Ahmad *et al.* (2016), at an alpha level of 0.05. This can be attributed to the relatively small group of validation datasets provided in the FCPS (i.e., 10 datasets).



Note: p -value: <0.001 ***, <0.01 ** , <0.05 *

Figure 2. Average Rand index for FCPS datasets



Note: p -value: <0.001 ***, <0.01 ** , <0.05 *

Figure 3. Proportion of results with Rand index of 1 for FCPS datasets (i.e., perfect clustering)

Similarly, Figure 3 shows the proportion of the FCPS datasets that achieved a clustering result with a Rand index of 1 (i.e., perfect clustering) for each of the given methods. The results reveal that the proposed method was able to obtain a Rand index of 1 in 6 out of the 10 FCPS datasets. The results from the

Wilcoxon tests indicate that these results are statistically significantly greater than the results of the methods proposed by Ultsch and Lötsch (2017), Cheng and Church (2000), Lee and Seung (2001), and Sakellariou *et al.* (2012). Even though the results indicate the proposed method correctly clusters the largest percentages of datasets (i.e., 6/10), there is not enough evidence to conclude that this proportion is statically significantly greater than the ones from the other methods benchmarked, at an alpha level of 0.05. Nevertheless, these results provide evidence that the method presented in this work is able to identify true clusters in a wider range of datasets with different underlying structures.

5.2. Genomic dataset Benchmark results

Figure 4 presents the Rand index obtained on the Golub *et al.* (1999) dataset [57] by the method proposed in this work and the benchmark methods that do not require providing the number of clusters *a priori*. Fig. 4 indicates that the proposed method performed better than the methods presented by Clifford *et al.* (2011), Cheng and Church (2000), and Sakellariou *et al.* (2012).

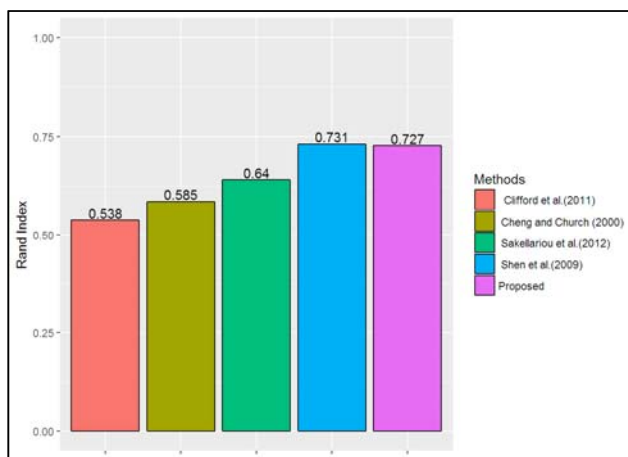
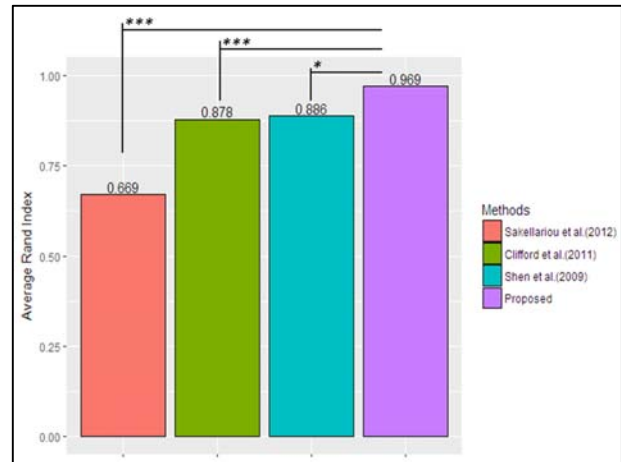


Figure 4. Rand index for Leukemia dataset

Figure 5 shows the average Rand index obtained with the MS dataset and the 10-fold cross-validation approach by the proposed and benchmark methods. The iterative greedy search Biclustering algorithm proposed by Cheng and Church (2000) was not able to find any cluster structure in the MS dataset; hence it was not included in this plot. The plot shows that on average the proposed method outperformed the other methods, with an average Rand index of 0.969. This is statistically significantly greater than the values obtained with the other methods benchmarked. Moreover, the average Rand index obtained by the proposed method was not significantly different than an average Rand index of 1 (t -value: -1.963, p -value=0.0812), at an alpha

level of 0.05. This reveals that on average the proposed method found a perfect match between the clusters of patients obtained with the complete MS dataset and the cross-validation subsets.



Note: p -value: <0.001***, <0.01**, <0.05*

Figure 5. Average Rand index for MS dataset

Table 4 shows the confusions matrix of the clusters obtained with the proposed method when implementing the 10-fold cross-validation approach. The table indicates that the proposed method was able to group 96.33% of the patients' in the same clusters when both the complete dataset and the different data subsets were used (i.e., accuracy of 0.96), which is in line with the average Rand index of 0.969 shown in Fig. 5. The Rand index and confusion matrix results indicate that the proposed method identified a similar cluster structure even with different subsets of the MS dataset. This indicates that the proposed method was not overfitting the dataset. Furthermore, it provides evidence that supports that the method was able to identify clusters due to the underlying structure of the data and not just due to random change.

Table 4. MS dataset 10-fold cross-validation confusion matrix

		Complete Dataset	
		Cluster 1	Cluster 2
Data Subsets	Cluster 1	172	0
	Cluster 2	7	12

The results from the pathway analysis on the set of statistically significant different SNPs between the MS patient clusters are shown in Fig. 6. The cluster-defining SNPs show significantly more interactions than expected among a random sampling of genes. Out of 515 genes, 1,463 interactions were found, with only 942 expected by chance (p -value: 1.04e-10), among a background set of 4,938 genes present on the

Moreover, the method is capable of automatically identifying the number of clusters by implementing an internal validity metric. Similarly, the method takes advantage of the degree of linkage disequilibrium between SNPs by pruning correlated nearby SNPs, as well as implementing a post-clustering gene pathways analysis.

The method is tested with clustering validation datasets previously used in the literature. The benchmark results reveal that proposed method provides, on average, the greatest performance (i.e., average Rand index 0.852). Moreover, results indicate that it was able to obtain cluster results with a Rand index of 1 (i.e., perfect clustering) in 6 out of the 10 Fundamental Clustering Problem Suite datasets. Similarly, the method is applied to a dataset of 38 patients with leukemia, and subsequently to a dataset of 191 Multiple Sclerosis (MS) patients. The results indicate that the method is able to identify genetically distinct patient clusters without the need to select the number of clusters or any input parameter *a priori*. Moreover, the cross-validation results indicate that the method presented in this work outperformed the other methods found in the literature when it comes to data overfitting, since the average Rand index obtained was significantly greater than the benchmarked methods and not significantly different than 1. This performance was maintained even after the removal of potentially related patients from the dataset. This indicates that the method was identifying clusters due to the underlying structure of the data and avoided overfitting the dataset. The identification of distinct genetic subtypes of patients demonstrates the potential applicability of this process to advance personalized medicine of complex diseases with heritable components, especially autoimmune disorders.

When applied to genomic data, the method also shows value as a feature reduction strategy. Out of over 25,482 exonic SNPs and 191 patient samples, the clustering of patients yielded a set of SNPs which significantly vary between clusters. These variants represent 515 genes, several of which are known to be involved in MS (CD69, CCRX5, IL-13, STAT3) and cell adhesion (ICAM1, LAMB4). The fact that many highlighted genes are components of the immune system is not surprising due to the nature of the Immunochip assay, but the enrichment of leukocyte-specific genes is evidence that the method can result in functionally relevant feature sets, even without class labels. Notably, 57 genes representing over 10% of the network are involved in cytokine receptor processes. This is greater than expected from random chance, as cytokine receptors constitute a small percentage of all Immunochip genes. The evidence

presented in this work alone is insufficient to define genetic subtypes of MS, but the specific SNP set reaching significance may be a valuable resource in experimental studies examining immune cell dynamics and genetics. For example, the hypothesis that these clusters represent different subtypes of MS, can be tested by evaluating clinical criteria such as image results and disease progression, as well as quantitative cytokine profiling and gene expression studies for each cluster, compared against random groupings of patients.

This work demonstrates an iterative unsupervised machine learning method which identifies significant patient clusters within a genomic dataset. Future research should explore the medical significance of the findings shown in this work. Similarly, the method from this work should be implemented in studies collecting SNP array and gene expression microarray data from additional disease cohorts to explore its potential benefits. Further investigation can extend beyond pathway analysis to evaluate these clusters for clinical significance of genetically related characteristics such as age of onset, disease course, heritability, and response to treatment. Once links are drawn between clusters and clinically relevant outcomes, the Immunochip can be used to classify high-risk and newly diagnosed chronic disease patients into clusters with predictive value.

Acknowledgments

The authors acknowledge the NSF I/UCRC Center for Healthcare Organization Transformation (CHOT), NSF I/UCRC grant #1624727, and the Institute for Personalized Medicine at the Pennsylvania State University. Additionally, the authors would like to acknowledge Dr. James R. Broach from the Institute for Personalized Medicine at the Pennsylvania State University, for his valuable contributions. Any opinions, findings, or conclusions found in this paper are those of the authors and do not necessarily reflect the views of the sponsors.

References

- [1] M.K.K. Leung, A. Delong, B. Alipanahi, B.J. Frey, Machine Learning in Genomic Medicine: A Review of Computational Problems and Data Sets, Proceedings of the IEEE. 104 (2016) 176–197. doi:10.1109/JPROC.2015.2494198.
- [2] M.W. Libbrecht, W.S. Noble, Machine learning in genetics and genomics, Nature Reviews. Genetics. 16 (2015) 321–332. doi:10.1038/nrg3920.Machine.
- [3] R. Upstill-Goddard, D. Eccles, J. Fliege, A. Collins, Machine learning approaches for the discovery of gene-gene interactions in disease data, Briefings in

- Bioinformatics. 14 (2013) 251–260. doi:10.1093/bib/bbs024.
- [4] K.Y. Yip, C. Cheng, M. Gerstein, Machine learning and genome annotation: a match meant to be?, *Genome Biology*. 14 (2013) 205. doi:10.1186/gb-2013-14-5-205.
- [5] C.J. Ross, F. Towfic, J. Shankar, D. Laifenfeld, M. Thoma, M. Davis, B. Weiner, R. Kusko, B. Zeskind, V. Knappertz, I. Grossman, M.R. Hayden, A pharmacogenetic signature of high response to Copaxone in late-phase clinical-trial cohorts of multiple sclerosis, *Genome Medicine*. 9 (2017). doi:10.1186/s13073-017-0436-y.
- [6] O. Kulakova, E. Tsareva, D. Lvovs, A. Favorov, A. Boyko, O. Favorova, Comparative pharmacogenetics of multiple sclerosis: INF-B versus glatiramer acetate, *Pharmacogenomics*. 15 (2014) 679–85.
- [7] W. Xu, L. Zhang, Y. Lu, SD-MSAEs: Promoter recognition in human genome based on deep feature extraction, *Journal of Biomedical Informatics*. 61 (2016) 55–62. doi:10.1016/j.jbi.2016.03.018.
- [8] Y. Zhao, B.C. Healy, D. Rotstein, C.R.G. Guttmann, R. Bakshi, H.L. Weiner, C.E. Brodley, T. Chitnis, Exploration of machine learning techniques in predicting multiple sclerosis disease course., *PLoS One*. 12 (2017) e0174866. doi:10.1371/journal.pone.0174866.
- [9] B.A. Goldstein, A.E. Hubbard, A. Cutler, L.F. Barcellos, An application of Random Forests to a genome-wide association dataset: Methodological considerations & new findings, *BMC Genetics*. 11 (2010) 49. doi:10.1186/1471-2156-11-49.
- [10] P.K. Kimes, Y. Liu, D. Neil Hayes, J.S. Marron, Statistical significance for hierarchical clustering, *Biometrics*. (2017). doi:10.1111/biom.12647.
- [11] K.K.-H. Farh, A. Marson, J. Zhu, M. Kleinewietfeld, W.J. Housley, S. Beik, N. Shores, H. Whitton, R.J.H. Ryan, A.A. Shishkin, M. Hatan, M.J. Carrasco-Alfonso, D. Mayer, C.J. Luckey, N.A. Patsopoulos, P.L. De Jager, V.K. Kuchroo, C.B. Epstein, M.J. Daly, D.A. Hafler, B.E. Bernstein, Genetic and epigenetic fine mapping of causal autoimmune disease variants, *Nature*. 518 (2015) 337–343. doi:10.1038/nature13835.
- [12] S. Lim, C.S. Tucker, S. Kumara, An unsupervised machine learning model for discovering latent infectious diseases using social media data, *J Biomed Informat*. 66 (2017) 82–94.
- [13] R. Xu, D.C. Wunsch, Clustering algorithms in biomedical research: A review, *IEEE Reviews in Biomedical Engineering*. 3 (2010) 120–154. doi:10.1109/RBME.2010.2083647.
- [14] A. Prelic, S. Bleuler, P. Zimmermann, A. Wille, P. Bühlmann, W. Gruissem, E. Zitzler, A systematic comparison and evaluation of biclustering methods for gene expression data, *Bioinformatics*. 9 (2006) 1122–1129.
- [15] A.K. Jain, M.N. Murty, P.J. Flynn, Data clustering: a review, *ACM Computing Surveys*. 31 (1999) 264–323. doi:10.1145/331499.331504.
- [16] I.A. Pagnuco, Juan I. Pastorea, G. Abras, M. Brun, V.L. Ballarin, Analysis of genetic association using hierarchical clustering and cluster validation indices, *Genomics*. (2017) 4–11. doi:10.1016/j.pscychresns.2008.11.004.
- [17] T. Ahmad, N. Desai, F. Wilson, P. Schulte, A. Dunning, D. Jacoby, L. Allen, M. Fiuzat, J. Rogers, G.M. Felker, Clinical implications of cluster analysis-based classification of acute decompensated heart failure and correlation with bedside hemodynamic profiles, *PLoS One*. 11 (2016) e0145881.
- [18] K. Mei, J. Peng, L. Gao, N.N. Zheng, J. Fan, Hierarchical Classification of Large-Scale Patient Records for Automatic Treatment Stratification, *IEEE Journal of Biomedical and Health Informatics*. 19 (2015) 1234–1245. doi:10.1109/JBHI.2015.2414876.
- [19] J.S. Hamid, C. Meaney, N.S. Crowcroft, J. Granerod, J. Beyene, Cluster analysis for identifying sub-groups and selecting potential discriminatory variables in human encephalitis., *BMC Infectious Diseases*. 10 (2010) 364. doi:10.1186/1471-2334-10-364.
- [20] W. Redekop, D. Mladi, The faces of personalized medicine: a framework for understanding its meaning and scope, *Value in Health*. 6 (2013) S4–S9.
- [21] L. Tari, C. Baral, S. Kim, Fuzzy c-means clustering with prior biological knowledge, *Journal of Biomedical Informatics*. 42 (2009) 74–81. doi:10.1016/j.jbi.2008.05.009.
- [22] R. Bellazzi, B. Zupan, Towards knowledge-based gene expression data mining, *Journal of Biomedical Informatics*. 40 (2007) 787–802. doi:10.1016/j.jbi.2007.06.005.
- [23] C. Wiwie, J. Baumbach, R. Röttger, Comparing the performance of biomedical clustering methods, *Nature Methods*. 12 (2015) 1033–1038. doi:10.1038/nmeth.3583.
- [24] A. Ultsch, J. Lötsch, Machine-learned cluster identification in high-dimensional data, *Journal of Biomedical Informatics*. 66 (2017) 95–104. doi:10.1016/j.jbi.2016.12.011.
- [25] A. Ultsch, Clustering with SOM: U*C., in: *Proceedings of the 5th Workshop on Self-Organizing Maps, Paris, 2005*: pp. 75–82.
- [26] H. Clifford, F. Wessely, S. Pendurthi, R.D. Emes, Comparison of clustering methods for investigation of genome-wide methylation array data, *Frontiers in Genetics*. 2 (2011) 1–11. doi:10.3389/fgene.2011.00088.
- [27] P.J. Rousseeuw, Silhouettes: A graphical aid to the interpretation and validation of cluster analysis, *Journal of Computational and Applied Mathematics*. 20 (1987) 53–65. doi:10.1016/0377-0427(87)90125-7.
- [28] J.C. Bezdek, N.R. Pal, Some new indexes of cluster validity, *IEEE Transactions on Systems, Man, and Cybernetics, Part B: Cybernetics*. 28 (1998) 301–315. doi:10.1109/3477.678624.
- [29] A. Sakellariou, D. Sanoudou, G. Spyrou, Combining multiple hypothesis testing and affinity propagation clustering leads to accurate, robust and sample size independent classification on gene expression data, *BMC Bioinformatics*. 13 (2012) 270.
- [30] B.J. Frey, D. Dueck, Clustering by passing messages between data points, *Science*. 315 (2007) 972–976.
- [31] R. Shen, A.B. Olshen, M. Ladanvi, Integrative clustering of multiple genomic data types using a joint latent variable model with application to breast and lung cancer subtype analysis, *Bioinformatics*. 25 (2009) 2906–2912.
- [32] A.P. Dempster, N.M. Laird, D.B. Rubin, Maximum likelihood from incomplete data via the EM algorithm, *J Royal Stat Soc Series B*. (1977) 1–38.
- [33] C. Fraley, A.E. Raftery, How many clusters? Which clustering method? Answers via model-based cluster analysis., *The Comp Journal*. 41 (1998) 578–588.
- [34] S. Khakabimamaghani, M. Ester, Bayesian biclustering for patient stratification, *Biocomputing 2016: Proceedings of the Pacific Symposium*. (2016) 345–356.

- [35] D. Lee, H. Seung, Algorithms for non-negative matrix factorization, *Advances in Neural Information Processing Systems*. (2001) 556–562.
- [36] B. Pontes, R. Giráldez, J. Aguilar-Ruiz, Biclustering on expression data: A review, *Journal of Biomedical Informatics*. 57 (2015) 163–180.
- [37] Y. Cheng, G. Church, Biclustering of expression data, *Proceedings of the 8th International Conference on Intelligent Systems for Molecular Biology*, La Jolla, CA. (2000) 93–103.
- [38] N. Nguyen, R. Caruana, Consensus clusterings, in: *Proceedings - IEEE International Conference on Data Mining, ICDM, 2007*: pp. 607–612. doi:10.1109/ICDM.2007.73.
- [39] R. Suzuki, H. Shimodaira, pvclust: An R package for hierarchical clustering with p-values, *Bioinformatics*. 22 (2013) 1–7.
- [40] P.R. Bushel, H.K. Hamadeh, L. Bennett, J. Green, A. Ableson, S. Misener, C.A. Afshari, R.S. Paules, Computational selection of distinct class- and subclass-specific gene expression signatures, *Journal of Biomedical Informatics*. 35 (2002) 160–170. doi:10.1016/S1532-0464(02)00525-7.
- [41] R.J. Cabin, R.J. Mitchell, To Bonferroni or not to Bonferroni: when and how are the questions, *Bulletin of the Ecological Society of America*. 81 (2000) 246–248. doi:10.2307/20168454.
- [42] J.C. Bezdek, R. Ehrlich, FCM: The fuzzy c-means clustering algorithm, *Comp and Geosci*. 10 (1984) 191–203.
- [43] M. Ashburner, C.A. Ball, J.A. Blake, D. Botstein, H. Butler, J.M. Cherry, A.P. Davis, K. Dolinski, S.S. Dwight, J.T. Eppig, M.A. Harris, D.P. Hill, L. Issel-Tarver, A. Kasarskis, S. Lewis, J.C. Matese, J.E. Richardson, M. Ringwald, G.M. Rubin, G. Sherlock, Gene Ontology: tool for the unification of biology, *Nature Genetics*. 25 (2000) 25–29. doi:10.1038/75556.
- [44] A. Yazdani, A. Yazdani, A. Samiei, E. Boerwinkle, Generating a robust statistical causal structure over 13 cardiovascular disease risk factors using genomics data, *Journal of Biomedical Informatics*. 60 (2016) 114–119. doi:10.1016/j.jbi.2016.01.012.
- [45] S. Purcell, B. Neale, K. Todd-Brown, L. Thomas, M.A.R. Ferreira, D. Bender, J. Maller, P. Sklar, P.I.W. de Bakker, M.J. Daly, P.C. Sham, PLINK: A Tool Set for Whole-Genome Association and Population-Based Linkage Analyses, *The American Journal of Human Genetics*. 81 (2007) 559–575. doi:10.1086/519795.
- [46] G.P. Rédei, Kruskal-Wallis test, *Encyclopedia of Genetics, Genomics, Proteomics, and Informatics*. (2008) 1067–1068.
- [47] B.S. Everitt, S. Landau, M. Leese, D. Stahl, *Measurement of Proximity, Cluster Analysis*. (2011) 43–69. doi:10.1002/9780470977811.ch3.
- [48] M.S. Bazaraa, H.D. Sherali, C.M. Shetty, *Nonlinear programming: theory and algorithms*, John Wiley & Sons, Chicago, 2013.
- [49] Szklarczyk et al, STRING v10: protein-protein interaction networks, integrated over the tree of life, *Nucleic Acids Research*. D1 (2015) 447–452.
- [50] W.M. Rand, Objective criteria for the evaluation of clustering methods, *J Amer Stat Assoc*. 66 (1971) 846–50.
- [51] R. R Development Core Team, *R: A Language and Environment for Statistical Computing*, 1 (2011).
- [52] C. Fraley, A.E. Raftery, T.B. Murphy, L. Scrucca, mclust Version 4 for R: Normal Mixture Modeling for Model-Based Clustering, Classification, and Density Estimation, Technical Report No. 597, Dept of Statistics, University of Washington. (2012).
- [53] U. Bodenhofer, A. Kothmeier, S. Hochreiter, APCluster: an R package for affinity propagation clustering, *Bioinformatics*. 27 (2011) 2463–4.
- [54] M.C. Thrun, F. Lerch, J. Lotsch, A. Ultsch, Visualization and 3D printing of multivariate data of biomarkers, in: *Proceedings of International Conference in Central Europe on Computer Graphics, Visualization, and Computer Vision, Plzen*, 2016.
- [55] M.C. Thrun, *Projection based clustering through self-organization and swarm intelligence: combining cluster analysis with the visualization of high-dimensional data.*, Springer Fachmedien, Wiesbaden, Germany, 2018.
- [56] S. Kaiser, R. Santamaria, T. Khiamiakova, M. Sill, R. Theron, L. Quintales, F. Leisch, E. DeTroyer, biclust: BiCluster Algorithms, R Package Version 1. no. 1 (2015).
- [57] T. Golub, D. Slonim, P. Tamayo, C. Huard, M. Gaasenbeek, J. Mesirov, H. Coller, M. Loh, J. Downing, M. Caligiuri, C. Bloomfield, E. Lander, Molecular classification of cancer: class discovery and class prediction by gene expression monitoring, *Science*. 5439 (1999) 531–7.
- [58] S. Monti, P. Tamayo, J. Mesirov, T. Golub, Consensus clustering: a resampling-based method for class discovery and visualization of gene expression microarray data, *Machine Learning*. 52 (2003) 91–118.
- [59] A. Cortes, M.A. Brown, Promise and pitfalls of the Immunochip, *Arthritis Research & Therapy*. 13 (2011) 101.
- [60] D. Welter, J. MacArthur, J. Morales, T. Burdett, P. Hall, H. Junkins, A. Klemm, P. Flicek, T. Manolio, L. Hindorf, H. Parkinson, The NHGRI GWAS Catalog, a curated resource of SNP-trait associations, *Nucleic Acids Research*. 42 (2014) D1001–D1006. doi:10.1093/nar/gkt1229.
- [61] S. Kirkpatrick, C.D. Gelatt, M.P. Vecchi, Optimization by simulated annealing, *Science*. 220 (1983) 671–80.
- [62] C.E. López, D. Nembhard, Cooperative workforce planning heuristic with worker learning and forgetting and demand constraints, in: *IIE Annual Conference Proceedings*, 2017: pp. 380–85.
- [63] Y. Xiang, S. Gubian, B. Suomela, J. Hoeng, Generalized simulated annealing for global optimization: the GenSA Package, *R J*. 5 (2013) 13–28.
- [64] G.D. Bader, D. Betel, C.W.V. Hogue, BIND: the Biomolecular Interaction Network Database, *Nucleic Acids Research*. 31 (2003) 248–50.
- [65] A. Chatr-Aryamontri, R. Oughtred, L. Boucher, J. Rust, C. Chang, N.K. Kolas, L. O'Donnell, S. Oster, C. Theesfeld, A. Sellam, C. Stark, B.J. Breitkreutz, K. Dolinski, M. Tyers, The BioGRID interaction database: 2017 update, *Nucleic Acids Research*. (2016).
- [66] T.S.K. Prasad et al, Human Protein Reference Database - 2009 Update, *Nucleic Acids Research*. (2009) D767-72.
- [67] H. Hermjakob, L. Montecchi-Palazzi, C. Lewington et al, IntAct: an open source molecular interaction database, *Nucleic Acids Research*. (2004) D452-5.
- [68] L. Licata, L. Briganti, D. Peluso, L. Perfetto, M. Iannuccelli, E. Galeota, F. Sacco, A. Palma, A.P. Nardoza, E. Santonico, L. Castagnoli, G. Cesareni, MINT, the molecular interaction database: 2012 update, *Nucleic Acids Research*. (2012).

- [69] C.F. Schaefer, K. Anthony, S. Krupa et al, PID: the Pathway Interaction Database, *Nucleic Acids Research*. (2009) D674-9.
- [70] Kanehisa, M. Furumichi, M. Tanabe, Y. Sato, K. Morishima, KEGG: new perspectives on genomes, pathways, diseases, and drugs, *Nucleic Acids Research*. (2017) D353-61.
- [71] D. Nishimura, Biotech software and internet report, BioCarta, *Biotech Software & Internet Report: The Computer Software Journal for Scientist* 2(3). (2004). <https://doi.org/10.1089/152791601750294344>.
- [72] Fabregat et al, The reactome pathway knowledgebase, *Nucleic Acids Research*. D1 (2015) 481–487.
- [73] C. Larochelle, J. Alvarez, W. Just, How do immune cells overcome the blood-brain barrier in multiple sclerosis?, *FEBS Letters*. (2012).
- [74] N. Isobe, et. al., An Immunochip study of multiple sclerosis risk in African Americans, *Brain*. 138 (2015) 1518–30.